A water-soluble polysaccharide isolated from seeds of Cassia ovata

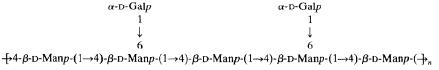
PRAMOD KUMAR*, VANDANA SINGH, UMESH CHANDRA MISHRA, AND PURNA CHANDRA GUPTA Department of Chemistry, University of Allahabad, Allahabad (India) (Received October 13th, 1988; accepted for publication, August 16th, 1989)

The plants of *Cassia* are regarded as medicinal and are rich sources of polysaccharides¹. We now report on the title polysaccharide.

The polysaccharide, isolated from dried, crushed and defatted seeds of *Cassia ovata* by extraction with aqueous 1% acetic acid and repeated precipitation from its solution therein with ethanol, had $[\alpha]_D^{25}$ +68° (water). Its homogeneity was verified by fractional precipitation, acetylation–deacetylation and zone electrophoresis. The polysaccharide was water soluble, had an ash content of 0.35%, and had negligible contents of methoxyl, acetyl groups, and uronic acid.

Acid hydrolysis of the polysaccharide gave D-galactose and D-mannose in the molar ratio 2:5. Graded acid hydrolysis liberated galactose, suggesting that these residues were present as end groups. The methylated polysaccharide had $[\alpha]_D^{25}+11^\circ$ (chloroform), and hydrolysis gave 2,3-di-O-methyl-D-mannose, 2,3,6-tri-O-methyl-D-mannose, and 2,3,4,6-tetra-O-methyl-D-galactose in the molar ratios 2:3:2. Periodate oxidation of the polysaccharide liberated 0.180 mol of formic acid (90 h), indicating 28.7% of end groups. The galactose and mannose residues were completely oxidised in 90 h. These results accord with those of the methylation study. Partial hydrolysis of the polysaccharide with 0.05M sulphuric acid at 100° for 12 h gave mannobiose $[\beta$ -D-Manp-(1 \rightarrow 4)-D-Manp], epimelibiose $[\alpha$ -D-Galp-(1 \rightarrow 6)-D-Manp], mannotriose $[\beta$ -D-Manp-(1 \rightarrow 4)-D-Manp], together with galactose and mannose.

These results indicate that the polysaccharide is a $(1\rightarrow 4)-\beta$ -D-mannan substituted at positions 6 by D-galactosyl groups. A possible repeating unit of the polysaccharide is shown in 1.



^{*}Kamla Nehru Institute of Physical and Social Sciences, Sultanpur (Avadh), India.

NOTE 385

EXPERIMENTAL

Solutions were concentrated under diminished pressure at 60– 62° . All residues were dried *in vacuo* over anhydrous calcium chloride. Melting points are uncorrected and $[\alpha]_D$ values are for equilibria. P.c. was carried out at room temperature with A, 1-butanol–ethanol–water (5:1:4); B, 1-butanol–ethanol–water (4:1:5); C, 1-butanol–2-propanol–water (11:6:3); D, ethyl acetate–pyridine–water (10:4:3); and E, ethyl acetate–pyridine–water (2:1:2); with detection using aniline hydrogenphthalate.

Isolation of the polysaccharide. — Dried crushed seeds were extracted successively with light petroleum and ethanol to defat and decolorise. The defatted and decolorised seeds were extracted with aq. 2% acetic acid, and the extract was added slowly with stirring to a large excess of ethanol. The crude polysaccharide was collected, washed with ethanol, dried and precipitated from solution in aq. 1% acetic acid with ethanol. The product (yield, \sim 3.2 g/100 g) had $[\alpha]_D^{25}$ +66° (c 1, water), and gave 0.35% of ash.

The homogeneity of the polysaccharide was tested by fractional precipitation from its aqueous solution with ethanol. Each fraction had $[\alpha]_D^{25}$ +68° (water) and on hydrolysis with M sulphuric acid at 100° for 20 h, gave D-galactose and D-mannose in the molar ratio 2:5. The polysaccharide was subjected to zone electrophoresis^{2,3} on Whatman No. 1 paper in borate buffer (pH 9.2) at 320 V and 3.7 mA for 6 h. The paper was cut into 31 equal segments and each was eluted with distilled water. The intensity of the characteristic yellow-orange colour developed in each eluate by adding aq. 8% phenol (1 mL) and conc. H₂SO₄ (6 mL) was measured in a Klett-Summerson photoelectric colorimeter (filter No. 50). A plot of absorbance against segment number showed a single sharp peak.

The polysaccharide was treated with sodium acetate-acetic anhydride⁴ and the resulting acetate had $[\alpha]_D^{28}$ +27.5° (c 1.2, chloroform). Deacetylation⁴ generated material with $[\alpha]_D^{28}$ +57.5° (c 1.3, water).

Investigation of the structure of the polysaccharide. — The purified polysaccharide was completely hydrolysed with M H_2SO_4 at 100° for 20 h. P.c. (solvent D) of the hydrolysate revealed galactose (R_F 0.15) and mannose (R_F 0.21). The absolute configurations were confirmed by the preparation of D-galactose phenylosazone, m.p. 164°, $[\alpha]_D^{30}$ +80° (c 1.5, water), and the isolation of D-mannose, m.p. 131°, $[\alpha]_D$ +14° (c 1.8, water).

The polysaccharide (300 mg), together with D-ribose (30 mg) as reference, was treated with M H_2SO_4 at 100° for 20 h. P.c. and quantification² of the components in the hydrolysate revealed the molar ratio of galactose and mannose to be 2:5.

The polysaccharide was hydrolysed⁵ with 25mm H_2SO_4 at 100° for 6 h. P.c. (solvent C) of the hydrolysate showed that the galactose was liberated first.

To a solution of the polysaccharide were added KCl and 0.25M sodium metaperiodate⁶. The amount of formic acid liberated was 0.230 mol/100 g (72 h), corresponding to 28.7% of end groups.

The polysaccharide was methylated (Haworth⁷ then Purdie⁸). The product, $[\alpha]_D^{25} + 11^\circ$ (c 1.2, chloroform), was hydrolysed⁶ with aq. 90% formic acid at 100° for 6 h, then with M H_2SO_4 for 14 h at 100°, and the products were fractionated on Whatman No. 3 paper (solvent A) to give the following compounds. 2,3,4,6-Tetra-O-methyl-D-galactose, m.p. 72–73°, $[\alpha]_D^{32} + 120^\circ$ (c 1, water); lit.⁹ m.p. 74°, $[\alpha]_D^{32} + 121^\circ$ (water). 2,3-Di-O-methyl-D-mannose, m.p. 107–108°, $[\alpha]_D^{25} - 16^\circ$ (c 1.5, water); lit.⁹ m.p. 106°, $[\alpha]_D^{25} - 15.8^\circ$; the anilide¹⁰ had m.p. 136°. 2,3,6-Tri-O-methyl-D-mannose, $[\alpha]_D^{25} - 11^\circ$ (water); lit.¹¹ $[\alpha]_D^{25} - 10^\circ$ (water); the hydrazide¹² had m.p. 121–131°.

The methylated polysaccharide together with D-glucose as reference was treated with M $\rm H_2SO_4$ at 100° for 18 h. The resulting methylated sugars were subjected to p.c. (solvent A) and their molar ratios were determined by alkaline hypoiodite¹¹. The molar ratios of the three methylated sugars were 2:2:3.

The polysaccharide was hydrolysed with 0.25M H_2SO_4 at 100° for 12 h. Preparative p.c. (solvents *D* and *E*) of the hydrolysate gave D-galactose, D-mannose, and the following oligosaccharides. Mannobiose [β-D-Manp-(1→4)-D-Manp], m.p. 203–205° (from ethanol), $[\alpha]_D^{25} - 9^\circ$ (*c* 1.2, water); lit.^{12,13} m.p. 193–210°, $[\alpha]_D^{25} - 5$ to -9° (water). Epimelibiose [α-D-Galp-(1→6)-D-Manp], m.p. 199°, $[\alpha]_D^{32} + 120.5^\circ$ (*c* 1.3, water); lit.¹⁴ m.p. 200°, $[\alpha]_D^{32} + 121^\circ$ (water). Mannotriose [β-D-Manp-(1→4)-β-D-Manp-(1→4)-D-Manp], m.p. 211–213° (from ethanol), $[\alpha]_D^{25} - 13^\circ$ (*c* 1.2, water); lit.^{12,13} m.p. 214–215°, $[\alpha]_D - 15$ to -26° (water). Galactosylmannobiose $[\alpha$ -D-Galp-(1→6)-β-D-Manp-(1→4)-D-Manp], m.p. 225–227°, $[\alpha]_D^{25} + 93^\circ$ (*c* 1.5, water); lit.¹⁵ m.p. 228–229°, $[\alpha]_D^{25} + 93.3^\circ$ (water).

REFERENCES

- 1 L. BRUGGAMAN, Tropical Plants and their Cultivation, Thames and Hudson, London, 1957, p. 148.
- 2 S. N. KHANNA AND P. C. GUPTA, Phytochemistry, 6 (1967) 605-609.
- 3 A. B. Foster, Adv. Carbohydr. Chem., 12 (1957) 86-90.
- 4 A. S. CEREZO, J. Org. Chem., 30 (1965) 924-927.
- 5 F. SMITH AND R. MONTGOMERY, Chemistry of Plant Gums and Mucilages, Reinhold, New York, 1959, pp. 134-135.
- 6 F. Brown, T. G. Halsall, E. L. Hirst, and J. K. N. Jones, J. Chem. Soc., (1948) 28-32.
- 7 W. N. HAWORTH, J. Chem. Soc., (1915) 107-110.
- 8 T. Purdie and J. C. Irvine, J. Chem. Soc., (1903) 1021–1026.
- 9 G. J. ROBERTSON, J. Chem. Soc., (1934) 330-336.
- 10 E. L. HIRST AND J. K. N. JONES, J. Chem. Soc., (1948) 1278-1280.
- 11 E. L. Hirst, L. Hough, and J. K. N. Jones, J. Chem. Soc., (1949) 928-932.
- 12 A. TYMINISKI AND T. E. TIMELL, J. Am. Chem. Soc., 82 (1960) 2823-2827.
- 13 G. O. ASPINALL, R. B. RASHBROOK, AND G. KESSLER, J. Chem. Soc., (1958) 215-222.
- 14 R. W. Bailey (Ed.), Oligosaccharides, Vol. 4, Pergamon, Oxford, 1965, pp. 97-101.